Identification of the SIM1 and ARNT DNA fragments from the yeast two-hybrid plasmid carried by the clone 06L22 by duplex PCR. Plasmid DNA was isolated from a liquid culture of the clone 06L22 by a QiaPrep (Hilden) procedure and the inserts contained within the plasmids were amplified by PCR using the primer pairs, 5'-TCG TAG ATC TTC GTC AGC AG-3' (SEQ ID No. 4) & 5'-GGA ATT AGC TTG GCT GCA GC-3' (SEQ ID No. 5) for the plasmid pBTM117c and 5'-CGA TGA TGA AGA TAC CCC AC-3' (SEQ ID No. 6) & 5'-GCA CAG TTG AAG TGA ACT TGC-3' (SEQ ID No. 7) for pGAD427. Lane 1

a2

Please replace the fifth full paragraph on page 51 with:

For the construction of pGAD427 a 1.2 kb *Dde* I fragment containing the *aphA* gene was isolated from pFG101u (Pansegrau et al., 1987) and was subcloned into the *Pvu* I site of the pGAD426 using the oligonucleotide adapters 5'- GTCGCGATC-3' and 5'-TAAGATCGCGACAT-3' (SEQ ID No. 8). The plasmid pGAD426 was generated by insertion of a 1.2 kb *Eco* RV *CYH2* gene fragment, which was isolated from the pAS2-1 (Clonetech) into the *Pvu* II site of pGAD425 (Han and Collicelli, 1995).

a3

Please replace the fourth full paragraph on page 90 with:

Plasmids carrying teh mammalian readout systems named pG5E1bEGFPneo, pG5E1bEGFPhyg or pG5E1bEGFPpur are used. These plasmids contain the TATA element of the adenoviral E1b gene and five tandem copies of the GAL4 responsive element UAS_G (5′ CGGAGTACTGTCC TGCG 3' (SEQ ID No. 9)) (Sadowski, I., Ma, J., Treizenberg, S. and Ptashne, M. (1988), Nature 335: 559-560) positioned immediately upstream of the coding sequence for the enhanced green fluorescent protein (EGFP; Yang, T.T., Cheng, L. and Kain, S.R. (1996), Nucl. Acids Res. 24 (22): 4592-4593). These reporter plasmids are generated by replacing the coding sequence for CAT in G5E1bCAT (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962) by the EGFP coding sequence and introducing either a neomycin, hygromycin or puromycin resistance marker gene (neo^r, hyg^r or pur^r) using standard subcloning procedures.

a4

Please replace the first full paragraph on page 91 with:

The plasmids pVP-Nconeo and pVP-Ncohyg are derived from pVP-Nco vector (Tsan, J., Wang, Z., Jin, Y., Hwang, L., Bash, R.O., Baer, R. The Yeast Two-Hybrid System, edn 1. Edited by Bartel, P.L., Fileds, S. New York: Oxford University Press (1997): 217-232) by insertion of either a neo^r or hyg^r marker gene using standard subcloning procedures. pVP-Nco in turn is an improved version of the pNLVP16 plasmid, which was constructed for the expression of herpes simplex virus protein VP16-fusion proteins in mammalian cells (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962). A polylinker sequence is preceded by an artificial reading frame including the eleven amino-terminal residues of Gal4p (MKLLSSIEQAC (SEQ ID No. 10)), a nuclear localization signal from the SV40 large T antigen (PKKKRKVD (SEQ ID No. 11)) and the acidic transactivation domain (amino acids 411-456) of the VP16 protein. Alternatively, the hybrid reading frames comprising Gal4 (1-147) and individual sequences of a cDNA library are subcloned into pLXSN or any other similar retroviral vector to allow packaging cell line-aided infection of target cells.

Please replace the table on page 101 with:

Oligonucleotide	**Sequence (5'-3')**
a sense	TCGAGTCGACGCGGCCGCTAA (SEQ ID No. 12)
A antisense	GGCCTTAGCGGCCGCGTCGAC (SEQ ID No. 13)
b sense	TCGAGGTCGACGCGGCCGCAGTAA (SEQ ID No. 14)
B antisense	GGCCTTACTGCGGCCGCGTCGACC (SEQ ID No. 15)
c sense	TCGAGAGTCGACGCGGCCGCTTAA (SEQ ID No. 16)
c antisense	GGCCTTAAGCGGCCGCGTCGACTC (SEQ ID No. 17)

Please enter the sequence listing filed herewith.

The replacement paragraphs presented above incorporate changes as indicated by the marked-up versions below.

Polylinkers used within the multiple cloning site to provide expression of the genetic fragment in one of the three reading frames. (SEQ ID Nos. 1-3)

Identification of the SIM1 and ARNT DNA fragments from the yeast two-hybrid plasmid carried by the clone 06L22 by duplex PCR. Plasmid DNA was isolated from a liquid culture of the clone 06L22 by a QiaPrep (Hilden) procedure and the inserts contained within the plasmids were amplified by PCR using the primer pairs, 5'-TCG TAG ATC TTC GTC AGC AG-3' (SEQ ID No. 4) & 5'-GGA ATT AGC TTG GCT GCA GC-3' (SEQ ID No. 5) for the plasmid pBTM117c and 5'-CGA TGA TGA AGA TAC CCC AC-3' (SEQ ID No. 6) & 5'-GCA CAG TTG AAG TGA ACT TGC-3' (SEQ ID No. 7) for pGAD427. Lane 1

For the construction of pGAD427 a 1.2 kb *Dde* I fragment containing the *aphA* gene was isolated from pFG101u (Pansegrau et al., 1987) and was subcloned into the *Pvu* I site of the pGAD426 using the oligonucleotide adapters 5'- GTCGCGATC-3' and 5'-TAAGATCGCGACAT-3' (SEQ ID No. 8). The plasmid pGAD426 was generated by insertion of a 1.2 kb *Eco* RV *CYH2* gene fragment, which was isolated from the pAS2-1 (Clonetech) into the *Pvu* II site of pGAD425 (Han and Collicelli, 1995).

Plasmids carrying teh mammalian readout systems named pG5E1bEGFPneo, pG5E1bEGFPhyg or pG5E1bEGFPpur are used. These plasmids contain the TATA element of the adenoviral E1b gene and five tandem copies of the GAL4 responsive element UAS_G (5′ CGGAGTACTGTCC TGCG 3' (SEQ ID No. 9)) (Sadowski, I., Ma, J., Treizenberg, S. and Ptashne, M. (1988), Nature 335: 559-560) positioned immediately upstream of the coding sequence for the enhanced green fluorescent protein (EGFP; Yang, T.T., Cheng, L. and Kain, S.R. (1996), Nucl. Acids Res. 24 (22): 4592-4593). These reporter plasmids are generated by replacing the coding sequence for CAT in G5E1bCAT (Dang, C.V., Barrett, J., Villa-Garcia, M., Resar, L.M.S., Kato, G.J. and Fearon, E.R. (1991), Mol. Cell. Biol. 11: 954-962) by the EGFP coding sequence and introducing either a neomycin, hygromycin or puromycin resistance marker gene (neo^r, hyg^r or pur^r) using standard subcloning procedures.